

INTENDED USE

QC1™ Parasitology Slides are designed for the quality control of parasitology staining procedures. They are intended to validate both the quality of the staining reagents and the technique of the procedure.

SUMMARY

The microscopic identification of the intestinal protozoa is based upon observing the morphology of stained organisms. Staining facilitates identification by differentiating internal structures and separating organisms from background material and artifacts. A number of simple staining techniques have been developed which are satisfactory for diagnostic work. One of the most commonly used is the Wheatley's Trichrome Stain procedure.

The QC1 Parasitology Slides contain parasite-positive and parasite-negative substrates necessary to produce the correct staining results when performing the Wheatley's Trichrome Stain procedure. The positive area consists of a smear containing fixed *Giardia lamblia* trophozoites; the negative area is a similar smear that does not contain parasites.

FOR IN VITRO DIAGNOSTIC USE ONLY

PRECAUTIONS

QC1 Parasitology Slides contain substrates that are applied as a smear. Handle the slides by the edges and do not touch the surface of the slide as this may damage the substrates.

STABILITY AND STORAGE

Store slides at room temperature (15-30°C). The slides are stable to the stated expiration date when stored at the required temperature.

PROCEDURE

Materials Provided: QC1 Parasitology Slides.

Materials Not Provided: Staining reagents, coverslips, mounting medium, microscope.

Wheatley's Trichrome Stain Procedure

Reagent	Timing
70% Ethanol (0003359)	3-5 minutes. Remove and drain off excess liquid by touching edge of slide to absorbent material.
70% Ethanol	3-5 minutes. Remove and drain off excess liquid by touching edge of slide to absorbent material.
Wheatley's Trichrome Stain (0003351)	10 minutes. Remove and drain off excess liquid by touching edge of slide to absorbent material.
90% Acid-Ethanol (0003350)	1-3 seconds. Remove and drain off excess liquid by touching edge of slide to absorbent material. NOTE: When stain begins to run, transfer immediately to the next step.
100% Ethanol (0003356)	5-10 seconds. Remove and drain off excess liquid by touching edge of slide to absorbent material.
100% Ethanol	3 minutes. Remove and drain off excess liquid by touching edge of slide to absorbent material.
100% Ethanol	3 minutes. Remove and drain off excess liquid by touching edge of slide to absorbent material.
PRO-CLEAR™ (0003336) or Xylene (0003342)	10 minutes. Remove and drain off excess liquid by touching edge of slide to absorbent material and coverslip with mounting fluid.
PRO-CLEAR or Xylene	10 minutes. Remove and drain off excess liquid by touching edge of slide to absorbent material and coverslip with mounting fluid.

NOTE: Staining time can vary depending on the intensity desired for the final stain result.

NOTE: The Xylene substitute AmeriClear® does not yield appropriate stain results and should not be used.

EXPECTED RESULTS

Positive Control with Wheatley's Trichrome Stain:

The cytoplasm of the *Giardia lamblia* trophozoites stains blue-green and may be tinted with purple. Nuclei and internal structures will stain red/purple. Background vegetative material, bacteria and artifacts will stain shades of green, blue, purple or red.

Negative Control with Wheatley's Trichrome Stain:

Vegetative material, bacteria and other artifacts will stain shades of green, blue, purple or red.

LIMITATIONS OF PROCEDURES

The Wheatley's Trichrome Stain method is usually a trouble-free procedure when followed as directed. In some instances, due to the nature of the staining reagents and slide fixatives, microorganisms may migrate during the staining procedure onto the negative area of the slide. Any other problems that may occur will usually be one of the following:

Problem: Poor contrast of the chromatin material.

Cause: Over-decolorizing.

Solution: Decolorizing requires only a very brief contact, followed by an immediate dip in ethanol.

Problem: Stained preparation is "cloudy" with poor contrast of cellular detail.

Cause: Carryover of solutions from one step to another. Excessive staining (more than 30-40 slides) will weaken or dilute the stain.

Solution: Change all solutions regularly to avoid staining clarity problems

BIBLIOGRAPHY

1. Lennette, EH, et al. Manual of Clinical Microbiology. Third Edition. American Society for Microbiology. 1980. pp. 684-685.
2. Melvin, DM and Brooke, MM. Laboratory Procedures for the Diagnosis of Intestinal Parasites. 3rd Edition. US Department of Health and Human Services Pub. No. (CDC) 1982. 82-8282.
3. Wheatley, WB. A Rapid Staining Procedure for Intestinal Amoeba and Flagellates. American Journal of Clinical Pathology. 1951. Vol. 21:990-991.

CONTACT

CalibreScientific US, Inc. offers a complete line of reagents, stains, and QC1™ Quality Control Slides for AFB, Parasitology, Bacteriology, and Mycology processing, as well as O&P collection systems and concentration devices for Parasitology. For Technical Assistance, email Technical@AlphaTecSystems.com, and for Customer Service, email Sales@AlphaTecSystems.com, or call either [+1] 800.221.6058 (USA) or [+1] 360.260.2779 between 8AM and 4PM Monday through Friday, Pacific Time.

WARRANTY

This product is warranted by CalibreScientific US, Inc. to perform as described in the labeling and literature supplied. CalibreScientific US, Inc. disclaims any implied warranty or merchantability or fitness for any other purpose, and in no event shall CalibreScientific US, Inc. be liable for any consequential damages arising out of aforesaid express warranty.

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PRODUCT CODES

0003262 QC1 Parasitology Slides, 40/Box



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GLOSSARY OF SYMBOLS



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